Quality of life after treatment

612 VAGINAL RADIOFREQUENCY FOR THE TREATMENT OF GENITAL ATROPHY IN PATIENTS WITH ONCOLOGICAL HISTORY IN A PUBLIC HOSPITAL. LIFE AFTER CANCER
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Introduction/Background Menopausal symptoms can impact quality of life. The goal of this prospective research is the clinical improvement on vulvovaginal sphere with vulvovaginal radiofrequency in cancer survivors.

Methodology Between June 2019 and February 2020 we apply vulvovaginal radiofrequency to 11 menopausal patients unresponsive to standard treatments. Symptoms are checked 6 months later.

Requirements Benign cytology and normal examination Non active vaginal infection Stable oncological process for at least 5 years

Materials Monopolar vulvovaginal radiofrequency generator Dermatology Life Quality Index (DLQI) before and after (figure 1) Record of the mean main symptom and photo shooting.

Method Specific and approved by Hospital protocol and Informed consent Number of sessions depends on response Maximum power applied: 3000 ms in two rounds in vulva and vagina Clinical control at 3, 6 and 12 months

Results The mean age is 55 years with ages between 43 and 71. Natural or medical mean menopause age is 48.87 ± 8.17 years.

Clinical history of patients: 63.6% (n=7) breast cancer, 27.3% (n=3) early stage endometrial cancer and 9.1% (n=1) of benign metastasizing leiomyomatosis.

When consulted, 63.6% (n=7) of the patients complained mainly of dyspareunia, 18.2% (n=2) of itching and 18.2% (n=2) of dryness. The average time of previous treatment had been 13.3 months. 54.5% (n=6) had received treatment with moisturizers, 36.3% (n=4) with steroids and 9.1% (n=1) did not tolerate any topical treatment. Patients with a history of endometrial cancer receive radiofrequency exclusively in external genitalia.

The average power used is 2491 ms (1700–3000) They have received radiofrequency every 25.93 days with an average of 6 sessions per patient.

Qualitative evaluations According to the DLQI scale, patients presented symptoms before/after the treatment

- No effect on patient’s life: (n=0)/36.4% (n=4)
- Small effect: 45.4% (n=5)/54.5% (n=6)
- Moderate effect: 27.3% (n=3)/9.1% (n=1)
- Very large effect: 27.3% (n=3)/0% (n=0)

Of the 11 patients, not all of them have been followed for a year, so the assessment of their condition is presented after 6 months. In these the DLQI scale has varied clinically > 4 in 6 of them. In those that have not, however, the clinical range has changed in 8 of them (figure 2).

The improvement in quality of life is significant in this group (p < 0.008, Wilcoxon signed rank test) until treatment is completed and all patients are followed.

- Subjective evaluation:
  They show improvement after 1.5 sessions. The first thing is an increase in hydration and a decrease in itching. No burns, short- or long-term discomfort have been reported and treatment is well tolerated by 100% of patients. Immediately, 18.2% (n=2) of the patients showed slight discomfort but it disappeared spontaneously (figures 1 and 2).

Conclusion While we wait more cases and more time for their evolution, radiofrequency is presented to us as a good alternative for genital atrophy in those patients who are symptomatic, who do not respond to usual therapies and in whom treatment with local oestrogens may not be ideal. Well tolerated and with good clinical response.

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Translational research

428 GENOMIC INSTABILITY METRIC CONCORDANCE BETWEEN ONCOSCAN™, CYTOSNP AND AN FDA-APPROVED HRD TEST
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Introduction/Background Various biomarkers have been investigated to identify patients likely to respond to PARP inhibition. PARP inhibitor olaparib plus bevacizumab is approved by the US FDA as maintenance therapy for homologous recombination deficiency (HRD)–positive advanced ovarian cancer; the FDA contemporaneously approved a commercial assay as a companion diagnostic for HRD assessment that includes a genomic instability biomarker. Other genomic platforms
measuring HRD are available or in development, including single-nucleotide polymorphism (SNP) genotyping arrays designed to measure tumour-related copy number changes. We evaluated the performance of OncoScan™ (ThermoFisher) and Infinium CytoSNP-850K (CytoSNP; Illumina) for assessing HRD genomic instability.

**Methodology**
DNA extracted from pretreatment archival tumour samples (N=126 across 20 indications) was evaluated with Oncoscan™, CytoSNP and an FDA-approved HRD test. ASCAT (v2.5.1), using log R ratio and B-allele frequency of autosomal markers with GC wave correction, was used to evaluate copy number variation (CNV) and loss of heterozygosity (LOH). The genomic metrics were further generated with default parameters using previously reported algorithms1 for LOH,2 number of telomeric-allelic imbalance (NTAI)3 and large-scale state transition (LST)4; the aggregate HRD metric was the sum of the three components. The association between genomic metrics (with BRCA deleterious alterations) and an FDA-approved HRD test metric was calculated using AUROC. Correlations among continuous metrics were assessed using Spearman rank correlation coefficients.

**Results**
CNV segmentation and genomic metrics were successfully calculated for 120 (Oncoscan™), 106 (CytoSNP) and 126 (FDA-approved test) samples. Assessed by SNP genotyping arrays, the genomic metric as a continuous variable demonstrated good association with deleterious BRCA alterations (AUROC of HRD: Oncoscan™, 0.87; CytoSNP, 0.75) (table 1) and the FDA-approved test at cutoff 42 (AUROC of HRD: Oncoscan™, 0.92; CytoSNP, 0.91) (table 2). The genomic metric as a continuous variable showed good correlation with the FDA-approved HRD test metric (Spearman correlation of HRD: Oncoscan™, 0.82; CytoSNP, 0.81). The Spearman correlation of genomic metrics with the FDA-approved HRD test metric was 0.68 (LOH), 0.76 (TAI), 0.78 (LST) and 0.82 (HRD) for Oncoscan™ and 0.59 (LOH), 0.77 (TAI), 0.82 (LST) and 0.81 (HRD) for CytoSNP.

**Conclusion**
HRD as a continuous variable assessed by SNP genotyping arrays showed good correlation with an FDA-approved HRD test metric; SNP assays may potentially be able to identify most HRD-positive tumours if appropriate clinically relevant cutoffs can be determined.

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CIRCULATING TUMOUR CELLS IN BREAST AND OVARIAN CANCER: SIZE-BASED ISOLATION AND EX Vivo EXPANSION

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Introduction/Background Circulating tumour cells (CTCs) play a crucial role in cancer dissemination and cellular extravasation leading to metastasis. There are only a limited number of CTCs per clinically/ethically allowed cancer patient’s blood draw and expanding this population of cells in vitro is crucial in order to provide a reliable number of cells to analyse CTC biology. CTCs can grow in a hypoxic environment and the activation of hypoxia-inducible factor (HIF-1α) results in increased cell survival and cellular proliferation, leading to cancer progression. Our aim was to optimise cell culture conditions using cobalt chloride (CoCl2) as a chemical inducer of hypoxia that would allow us to examine growth of cells in real time. Primary ovarian cancer cells would be used for the hypoxia optimisation and conditions adapted ovarian/breast CTC cultures in vitro.

Methodology Primary ovarian cancer cells were cultured in modified media supplemented with various concentrations of CoCl2 for HIF1α induction (50, 100, 150 and 200 uM). Cell viability and the expression of HIF-1α, PHH3, EpCAM and HER2 were examined in these cells using either ELISA, Immunoblotting or Immunofluorescence techniques. CTCs were isolated from breast and ovarian cancer patients using the ScreenCell® Cyto R device and cultured in specially modified media optimised for CTC culture supplemented with 20% FCS, growth factors and additives including: FGF-2, FGF-10, Nicotinamide, Y-27632, Primocin and CoCl2. EpCAM and HER2 were examined in cultured and expanded CTCs using Immunofluorescence techniques.

Results HIF-1α expression was induced and cell proliferation and viability were maintained in the primary ovarian cancer cells at a concentration of 100 μM of CoCl2. Subsequently this concentration was used for the culturing of isolated CTCs. Using this condition, CTCs were successfully cultured and expanded for more than nine weeks. Based on the morphological and phenotypical characterisation, two phenotypes of CTCs were isolated from a breast cancer patient; epithelial-like expressed EpCAM and quasi-mesenchymal express HER2.

Conclusion We demonstrated the feasibility of culturing cancer patient blood derived CTCs under hypoxic conditions. We also demonstrated the presence of heterogenous CTC populations; classical epithelial-like CTCs and quasi-mesenchymal subtypes in a breast cancer patient and their corresponding molecular phenotypes. Our work also demonstrated the suitability of size-based isolation for this culturing approach.

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