

Conclusion Ngs can help classify rare diseases if the classical pathological diagnostics do not give a satisfying diagnosis. There are currently no clear treatment recommendations for STK11 adnexal tumors yet. International registries and solid clinical follow-up data are urgently needed to enhance our knowledge on these potentially aggressive tumors.

2022-RA-945-ESGO ANTITUMOUR ACTIVITY OF DOSTARLIMAB BY PD-L1 AND TUMOUR MUTATION BURDEN IN PATIENTS WITH MISMATCH REPAIR DEFICIENT AND PROFICIENT TUMOURS IN THE GARNET TRIAL

¹Susana Banerjee, ²Thierry André, ³Dominique Berton, ⁴Susan L Ellard, ⁵Begoña Jimenez, ⁶Vanessa Samouëlian, ⁷Lucy Gilbert, ⁸Valentina Boni, ⁹Xinwei Han, ¹⁰Grace Antony, ⁹Jennifer Veneris, ¹¹Ana Oaknin. ¹The Royal Marsden NHS Foundation Trust and Institute of Cancer Research, London, UK; ²Sorbonne University and Saint-Antoine Hospital, Paris, France; ³GINECO and Institut de Cancerologie de l'Ouest, Centre René Gauducheau, Saint-Herblain, France; ⁴BC Cancer-Kelowna, Kelowna, BC, Canada; ⁵Hospital Clínico Universitario Virgen de la Victoria, Malaga, Spain; ⁶Gynecologic Oncology Division, Centre Hospitalier de l'Université de Montréal (CHUM), Centre de Recherche du CHUM (CRCHUM) et Université de Montréal, Montreal, QC, Canada; ⁷Division of Gynecologic Oncology, McGill University Health Centre, Montreal, QC, Canada; ⁸*affiliation at time of study. Current affiliation NEXT Oncology, Universitario Quironsalud, *START Madrid CIOCC (Centro Integral Oncológico Clara Campal), Hospital Universitario HM Sanchinarro, Madrid, Spain; ⁹GSK, Waltham, MA; ¹⁰GSK, London, UK; ¹¹Vall d'Hebron University Hospital, Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain

10.1136/ijgc-2022-ESGO.878

Introduction/Background Dostarlimab is a programmed death 1 (PD-1) inhibitor approved as monotherapy in patients with mismatch repair deficient (dMMR) recurrent/advanced endometrial cancer (EC) that has progressed on or after platinum-based chemotherapy or solid tumours that have progressed on or after prior treatment, with no satisfactory alternative treatment options. We report a post hoc analysis of antitumour activity by PDL1 expression and tumour mutational burden (TMB) in patients with dMMR and MMR proficient (MMRp) solid tumours in the GARNET trial.

Q3W for 4 cycles, then 1000 mg IV Q6W until progression or discontinuation. TMB and PDL1 were exploratory biomarkers. TMB status was determined by FoundationOne test; TMB-high (TMB-H) was defined as ≥ 10 mutations/Mb. PDL1 expression was determined by combined positive score (CPS) by Ventana assay; PDL1-high (PDL1-H) was defined as CPS ≥ 1 . The study was not powered to assess antitumour activity within subgroups.

Results TMB-H and PDL1-H were common in dMMR solid tumours; PDL1-H was observed in 39.4% of MMRp EC tumours (table 1). Objective response rate (ORR) was higher in patients with TMB-H/PDL1-H tumours (55.6% for all cohorts, combined; Table). Safety for each cohort was previously reported.¹

Conclusion PDL1-H and TMB-H were frequently observed in the dMMR EC and non-EC cohorts, regardless of tumour type; PDL1-H was also prevalent in MMRp EC tumours. Although not a powered analysis, ORR by BICR per RECIST v1.1 was higher in patients with TMB-H and PDL1-H solid tumours. Across cohorts, dMMR status was predictive of response.

REFERENCE

1. Andre T, et al. *Ann Oncol* 2021;**32**(suppl 5):S829-S866. 991P.

2022-RA-968-ESGO IMMUNOTHERAPY RESPONSE MONITORING USING PERSONALIZED CIRCULATING TUMOR DNA ANALYSIS IN PATIENTS WITH RELAPSED GYNECOLOGIC MALIGNANCIES

¹Kayla Castaneda, ¹Jose Salvador Saldivar, ²Young Kwang Chae, ³Hemant Sindhu, ⁴Concepcion Diaz-Arrastia, ⁵Kassondra Grzankowski, ⁶Georges Azzi, ⁷Carly Bess Scalise, ⁷Ekaterrina Kalashnikova, ⁷Brittany Nicosia, ⁷Shilpa Tekula, ⁷Alexey Aleshin, ⁷Adam C ElNaggar. ¹Spero Women's Oncology Center, El Paso, TX; ²Northwestern University Feinberg School of Medicine, Chicago, IL; ³Southwest Cancer Care, Sierra Vista, AZ; ⁴Memorial Hermann, Houston, TX; ⁵Austin Cancer Center, Austin, TX; ⁶Holy Cross Health, Ft. Lauderdale, FL; ⁷Natera, Inc., Austin, TX

10.1136/ijgc-2022-ESGO.879

Introduction/Background Immunotherapy has transformed cancer care. Unfortunately, responses within gynecologic malignancies have been modest when compared to other disease sites. Biomarkers for early determination of treatment benefit are urgently needed to spare unnecessary toxicity and cost. We evaluated if circulating tumor DNA (ctDNA) dynamics enable early detection of progressive disease (PD) and treatment response in patients with recurrent, gynecologic malignancies receiving immunotherapy.

Methodology Longitudinal plasma samples (n=138) were collected from 25 patients with recurrent cervical (N=6), endometrial (N=12), or ovarian (N=7) cancers who received immunotherapy. A personalized, tumor-informed multiplex PCR assay (Signatera™ bespoke mPCR NGS assay) was used for the detection of ctDNA in plasma samples.

Results Pre-treatment samples were available for 9 patients (78% ctDNA detection rate) and all 25 patients had on-treatment samples (68% ctDNA detection rate). Serially ctDNA negative patients (3/15 with imaging) had no evidence of disease on-treatment. ctDNA clearance was observed in 3 (cervical, N=2; endometrial, N=1) of the remaining 12 patients and correlated with clinical benefit. ctDNA decreased in additional 2 patients, both with objective response, while all 7 patients with increased ctDNA had PD. Increased ctDNA

Abstract 2022-RA-945-ESGO Table 1

	A1 (dMMR EC) N=103	F (dMMR non-EC) N=106	A1+F (dMMR combined) N=209	A2 (MMRp EC) N=142	A1+A2+F (Total) N=351
Biomarker distribution, n (%)					
TMB					
High	85 (82.5)	79 (74.5)	164 (78.5)	9 (6.3)	173 (49.3)
Low	13 (12.6)	9 (8.5)	22 (10.5)	129 (90.8)	151 (43.0)
Unknown	5 (4.9)	18 (17.0)	23 (11.0)	4 (2.8)	27 (7.7)
PDL1					
High	56 (54.4)	52 (49.1)	108 (51.7)	56 (39.4)	164 (46.7)
Low	23 (22.3)	17 (16.0)	40 (19.1)	45 (31.7)	85 (24.2)
Unknown	24 (23.3)	37 (34.9)	61 (29.2)	41 (28.9)	102 (29.1)
ORR by BICR per RECIST v1.1, n/N (%; 95% CI)*					
Overall	46/103 (44.7, 34.9–54.8)	41/106 (38.7, 29.4–48.6)	87/209 (41.6, 34.9–48.6)	19/142 (13.4, 8.3–20.1)	—
TMB-L/PDL1-L (L/L)	1/5 (20.0, 0.5–71.6)	1/3 (33.3, 0.8–90.6)	2/8 (25.0, 3.2–65.1)	2/43 (4.7, 0.8–15.8)	4/51 (7.8, 2.2–18.9)
TMB-L/PDL1-H (L/H)	2/5 (40.0, 5.3–85.3)	1/2 (50.0, 1.3–98.7)	3/7 (42.9, 9.9–81.6)	7/50 (14.0, 5.8–26.7)	10/57 (17.5, 8.7–29.9)
TMB-H/PDL1-L (H/L)	5/17 (29.4, 10.3–56.0)	3/14 (21.4, 4.7–50.8)	8/31 (25.8, 11.9–44.6)	0/1 (0, 0–97.5)	8/32 (25.0, 11.5–43.4)
TMB-H/PDL1-H (H/H)	29/50 (58.0, 43.2–71.8)	22/43 (51.2, 35.5–66.7)	51/93 (54.8, 44.2–65.2)	4/6 (66.7, 22.3–95.7)	55/99 (55.6, 45.2–65.5)

*Only those patients with both known TMB status and known CPS were included in ORR calculations. BICR, blinded independent central review; CPS, combined positive score; dMMR, mismatch repair deficient; EC, endometrial cancer; H, high; L, low; MMRp, mismatch repair proficient; ORR, objective response rate; PDL1, programmed death ligand 1; RECIST v1.1, Response Evaluation Criteria in Solid Tumors version 1.1; TMB, tumour mutational burden.

Methodology GARNET (NCT02715284) is a phase 1, multi-centre, open-label, single-arm study of dostarlimab in patients with advanced/recurrent solid tumours. Three expansion cohorts enrolled patients based on MMR status: dMMR (A1) and MMRp (A2) advanced/recurrent EC, and dMMR non-EC solid tumours (F). Patients received dostarlimab 500 mg IV

predicted PD prior to imaging by an average of ~2.5 months (lead-time) and was significantly associated with worse progression-free survival (PFS) compared to patients with decreased ctDNA (HR=0.14, 95%CI: 0.03–0.60; $p<0.01$). TMB and MSI status (binary) were not predictive of response in univariate ($p=0.4$, $p=0.4$) analyses.

Conclusion ctDNA dynamics can accurately predict clinical benefit and allow for early prediction of PD in patients with advanced ovarian cancer receiving immunotherapy. Further study is warranted to evaluate the clinical utility of a personalized, tumor-informed ctDNA assay in patients with gynecologic malignancies undergoing systemic therapies.

2022-RA-1007-ESGO HIGH EXPRESSION OF FAP+ CANCER-ASSOCIATED FIBROBLASTS PREDICT POOR OUTCOME IN PATIENTS WITH HIGH-GRADE SEROUS OVARIAN CANCER WITH HIGH CD8-POSTIVE T-CELL INFILTRATION

¹Josefin Fernebro, ²Sara Corvigno, ²Arthur Mezheieusky, ³Josefin Severin Karlsson, ⁴Laura Martin De La Fuente, ⁴Sofia M Westbom-Fremer, ²Joseph Carlson, ⁵Paivi M Kannisto, ⁴Ingrid M Hedenfalk, ⁴Susanne M Malander, ²Arne Östman, ⁶Hanna Dahlstrand. ¹Medical Unit Pelvic Cancer, Theme Cancer, Karolinska University Hospital, Stockholm, Sweden; ²Department of Oncology-Pathology, Stockholm, Sweden; ³Department of Pathology and Cytology, Stockholm, Sweden; ⁴Division of Oncology and Pathology, Department of Clinical Sciences, Lund, Sweden; ⁵Department of Obstetrics and Gynecology, Lund, Sweden; ⁶Medical Unit Pelvic Cancer, Theme Cancer, Stockholm, Sweden

10.1136/ijgc-2022-ESGO.880

Introduction/Background Tumor biology studies have implied that fibroblasts act as negative regulators of immune cell function in cancer. We investigated the impact of FAP-positive cells in high-grade serous ovarian cancer (HGSC) in relation to CD8 expression.

Methodology A discovery cohort (N=113) of HGSC was subjected to immunohistochemistry (IHC) of FAP and CD8. Marker status was correlated with overall survival (OS) and progression-free survival (PFS). Findings were confirmed in a validation cohort (N=121) and in public available datasets (TCGA and GSE9891).

Results We confirmed previous findings that high density of CD8+ cells in HGSC is associated with longer OS compared to low density (HR 0.55; 95% CI 0.33–0.85; $p=0.008$). In the discovery cohort high intensity of FAP was associated with shorter median PFS in cases with high density of stromal CD8+ cells (11.4 versus 18.6 months) compared to low intensity of FAP ($p=0.007$). In contrast, high intensity of FAP was not associated with PFS in cases with low density of CD8+ cells. In the validation cohort, high intensity of FAP in the patients with high density of stromal CD8+ cells was associated with shorter OS compared to low intensity of FAP ($p=0.01$). This association was not seen in the cases with low density of CD8+ cells. The association between high FAP expression and poor outcome in the high density CD8+ group was confirmed in two independent gene-expression data sets, with a shorter PFS in the TCGA dataset and shorter PFS and OS in the GSE9891 dataset.

Conclusion The study shows a specific FAP positive fibroblast-subset of cases with poor prognosis restricted to a CD8 high density group of HGSC. Therapy targeting the immunosuppressive action of fibroblasts may be a tool to enhance the known positive prognostic effect of CD8-cells in ovarian cancer and may be explored in T-cell depended immune therapy.

2022-RA-1093-ESGO VALIDATION OF SELF-SAMPLING USE FOR A MULTIPLEXED BIOMARKER ASSAY FOR HPV AND DYSPLASIA DETECTION

Anna Sophie Skof, Eva-Maria Payrich, Maja Struck, Sarah Thies, Carola Schreckenberger, Jaled Sehoul, Andreas M Kaufmann. Klinik für Gynäkologie, Charité – Universitätsmedizin Berlin – corporate member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany

10.1136/ijgc-2022-ESGO.881

Introduction/Background The use of self-sampling in cervical cancer (CxCa) screening increases the number of participants and enables the inclusion of prior underscreened women in rural areas. For PCR and DNA-based testing self-sampling is as sensitive as physician-sampling. We compared self- and physician-sampling for analysis by QuantiGene-Molecular-Profilng-Histology assay (QG-MPH) to detect and grade cervical dysplasia in a triage setting.

Methodology Women with an equivocal screening result were recruited and a cervical sample (Cervex broom) was taken into ThinPrep/PreservCyt. Participants were asked to take a self-sample (Evalyn-Brush) and fill a questionnaire. Crude lysates were used for the QG-MPH assay. This multiplexed Luminex bead-based technology platform (QuantiGene 2.0) detects and quantifies the mRNA abundance of 18 Human Papillomavirus (HPV) genotype-specific oncogenes, reference genes and cellular biomarkers characterizing dysplasia stages, simultaneously. Formerly developed biomarker-based risk scores predict CIN2+, CIN3+, or CxCa.

Results Of 699 study participants, 601 performed self-sampling (85.9%). Invalid samples in QG-MPH was comparable between self- and physician-sampling with 16.1% and 14.9%, respectively. Of 132 histologically confirmed CIN3 lesions QG-MPH determined in the physician-taken sample 61.4% (n=81) as CIN3 or higher, 25.8% (n=34) as low-grade lesions, and 12.9% (n=17) were not evaluable. Of 109 self-samplers from CIN3 positive women QG-MPH determined 17.4% (n=19) as CIN3 or higher, 59.6% (n=65) as low-grade and 22.9% (n=25) were not evaluable. PCR-based HPV testing detected 78.2% of physician- and 74.9% of self-samples positive while QG-MPH 52.5% (n=315) and 32.3% (n=194), respectively. Concordance was 82.0% by PCR and 63.8% by QG-MPH.

Conclusion While cellularity of self-taken samples is sufficient for valid measurement by QG-MPH, less high-grade lesions and HPV-infections are detected. Optimization of cutoffs for the self-taken sample may improve the sensitivity. We hypothesize that ‘missed’ CIN3 by QG-MPH biomarker profiling may be non-progressor lesions. This will be investigated further.

2022-RA-1126-ESGO MESENCHYMAL PROGNOSTIC SIGNATURE IN OVARIAN CANCER

¹Katarzyna Marta Lisowska, ²Katarzyna Aleksandra Kujawa, ¹Joanna Patrycja Syrkis, ³Alexander Jorge Cortez, ¹Patrycja Jakubowska. ¹Center for Translational Research and Molecular Biology of Cancer, Maria Skłodowska-Curie National Research Institute of Oncology, Gliwice Branch, Gliwice, Poland; ²Center for Translational Research and Molecular Biology of Cancer, Maria Skłodowska – Curie National Research Institute of Oncology Gliwice Branch, Gliwice, Poland; ³Department of Biostatistics and Bioinformatics, Maria Skłodowska-Curie National Research Institute of Oncology, Gliwice Branch, Gliwice, Poland

10.1136/ijgc-2022-ESGO.882